

RNAseq: Normalization and differential expression I

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- Robinson, Oshlack. *A scaling normalization method for differential expression analysis of RNA-seq data*. Genome Biology. 2010
- Hardcastle, Kelly. *baySeq: Empirical Bayesian methods for identifying differential expression in sequence count data*. BMC Bioinformatics. 2010

Outline of the presentation

- 1 Introduction
- 2 Pairwise calibration (EdgeR)
- 3 Differential expression

Introduction

normalization:

- comparison of expression levels between genes within a sample (same scale)
- however technical effects introduce a bias in the comparison between samples
- \Rightarrow normalization is crucial before performing differential expression
- calibration method EdgeR takes advantage of within-sample comparability

differential expression:

- appropriate distribution for count data
- incorporate calibration parameters

Framework

$Y_{g,k}$... observed count for gene g in library k

$N_k = \sum_{g=1}^G Y_{g,k}$... total number of reads for library k

$\eta_{g,k}$... number of transcripts of gene g in library k

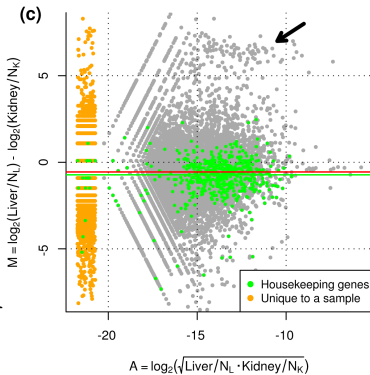
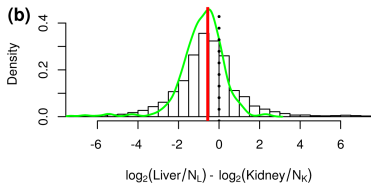
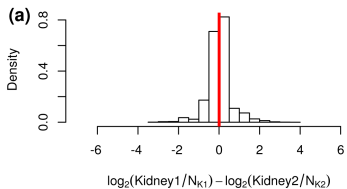
L_g ... length of gene g

$S_k = \sum_{g=1}^G \eta_{g,k} L_g$... total RNA output of sample k

$$E(Y_{g,k}) = \frac{\eta_{g,k} L_g}{S_k} N_k$$

- counts are a linear function of the number of transcripts
- library size calibration ($Y_{g,k}/N_k$) is appropriate for the comparison of replicates
- comparison of biologically different samples may be biased by varying RNA composition

kidney vs. liver dataset



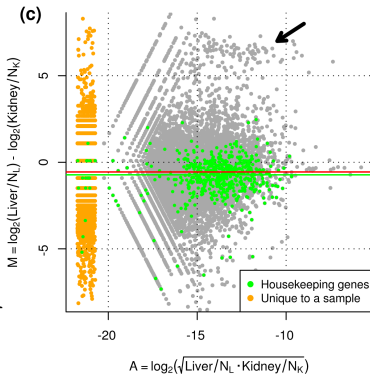
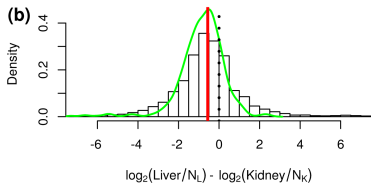
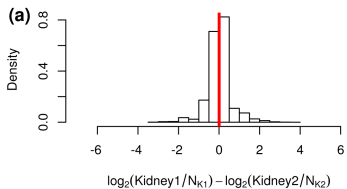
Trimmed mean of log-foldchange

- RNA production S_k of one sample cannot be determined directly
- estimation of relative differences of RNA production $f_k = S_k/S_r$ of a pair of samples (k, r)
- assumption: most genes are not differentially expressed
- \Rightarrow compute robust mean over log-foldchanges:
 - double filtering over both mean and difference of log-values
 - calculate a weighted mean over the log-foldchanges
 - \Rightarrow rescale factors $f_k = TMM_{(k,r)}$, where r is reference sample

$$\log_2 (TMM_{(k,r)}) = \frac{\sum_{g \in G^*} w_{g,(k,r)} (\log_2 (Y_{g,k}/N_k) - \log_2 (Y_{g,r}/N_r))}{\sum_{g \in G^*} w_{g,(k,r)}}$$

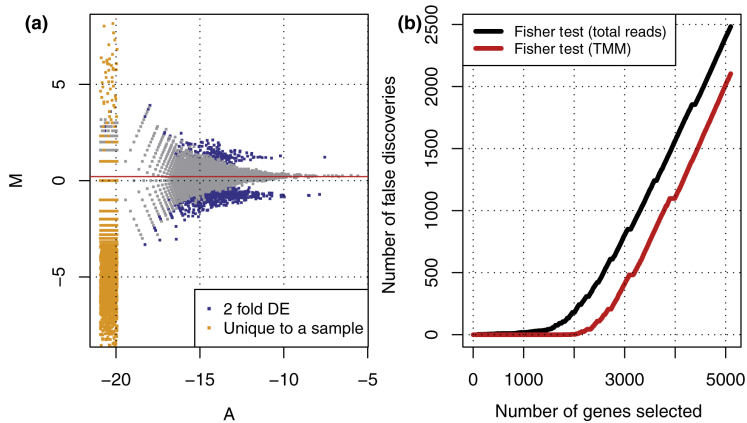
$$w_{g,(k,r)} = \left(\frac{1}{Y_{g,k}} - \frac{1}{N_k} + \frac{1}{Y_{g,r}} - \frac{1}{N_r} \right)^{-1}$$

kidney vs. liver dataset



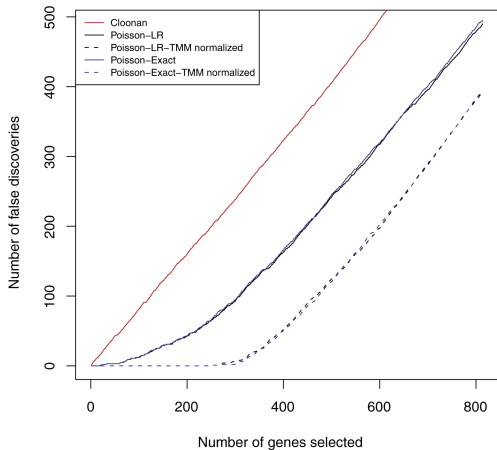
Simulation: pair of samples

simulated data sampled from poisson distribution



Simulation: replicates

Cloonan: log-transformation and quantile normalization



Differential expression

methods in use:

- DegSeq (normal distr.)
- EdgeR (negative binomial)
- DESeq (negative binomial, multiple groups)
- baySeq (negative binomial, multiple groups)
- Myrna (permutation based)

EdgeR

technical replicates: poisson distr.

biologically different samples: negative binomial distr.

$$Y \sim NB(p, m)$$

Y ... number of successes in a sequence of Bernoulli trials with probability p before r failures occur

alternative parametrization:

$q_{g,e}$... proportion of sequenced RNA of gene g for experimental group e

$$Y_{g,k,e} \sim NB(q_{g,e} N_k f_k, \phi_g)$$

$$E(Y_{g,k,e}) = \mu_{g,k,e} = q_{g,e} N_k f_k, \quad \text{Var}(Y_{g,k,e}) = \mu_{g,k,e} + \mu_{g,k,e}^2 \phi_g$$

- test if $q_{g,1}$ is significantly different from $q_{g,2}$
- dispersions ϕ_g are moderated towards a common dispersion

baySeq I

empirical Bayes approach to detect differential expression

$$D_g = \{Y_{g,k}, N_k, f_k\}_{k=1, \dots, K}$$

M ... user specified model

θ_M ... vector of parameters of model M

$$P(M|D_g) = \frac{P(D_g|M) P(M)}{P(D_g)}$$

calculate marginal likelihood:

$$P(D_g|M) = \int P(D_g|\theta_M, M) P(\theta_M|M) d\theta_M$$

baySeq II

$$P(D_g|M) = \int P(D_g|\theta_M, M) P(\theta_M|M) d\theta_M$$

- e.g. Poisson-Gamma conjugacy, however no such conjugacy with negative binomial data
- \Rightarrow define an empirical distribution on θ_M and estimate the marginal likelihood numerically

prior $P(M)$ is estimated by iteration:

$$P(M) = p_g, \quad p_g^* = P(M|D_g)$$

baySeq:

- applicable to complex experimental designs
- computationally intensive